

$\beta$ -ENDORPHIN: REPLACEMENT OF GLUTAMIC ACID  
IN POSITION 8 BY GLUTAMINE INCREASES ANALGESIC POTENCY  
AND OPIATE RECEPTOR-BINDING ACTIVITY

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SUMMARY

A  $\beta$ -endorphin analog with replacement of glutamic acid in position 8 by glutamine has been synthesized by modified procedures of the solid-phase method. The analgesic potency of the synthetic analog was increased to nearly three-fold with a concomitant increase of opiate receptor-binding activity in neuroblastoma x glioma hybrid cells. This is the first instance in which a replacement of a single amino acid causes an increase of analgesic potency of  $\beta$ -endorphin.

$\beta$ -Endorphin (1) is a potent analgesic agent by intracerebral (2) or intravenous (3) injections. So far, it has not been possible to obtain a synthetic  $\beta$ -EP analog with an analgesic potency more than twice that of the parent molecule (4). We report here a nearly 3-fold increase in the analgesic potency of  $\beta_h$ -EP (see Fig. 1) by replacement of a single amino acid glutamic acid in position 8 with a concomitant increase in

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Abbreviations:  $\beta_h$ -EP, human  $\beta$ -endorphin; Z, benzyloxycarbonyl; Bzl, benzyl; Boc, tert-butyloxycarbonyl; tlc, thin-layer chromatography

opiate receptor-binding activity. In addition, immunoreactivity of the synthetic analog decreases as revealed in the  $\beta_h$ -EP radioimmunoassay system.

#### MATERIALS AND METHODS

Synthesis of [Gln<sup>8</sup>]- $\beta_h$ -EP was carried out by modified procedures of the solid-phase method (5) as previously described (6). It was performed on Boc-Glu(Bzl)-brominated polymer (163 mg, 52  $\mu$ mole) with side-chain protection as follows: 2-Clz for Lys (7); 2-BrZ for Tyr-27 (8) and Z for Tyr-1 (9); Bzl for Thr and Ser. The completed protected peptide resin (459 mg) was treated with liquid HF (10) and the product isolated by gel filtration on Sephadex G-10 (0.5 N acetic acid) and chromatographed on carboxymethylcellulose (6). Final purification was effected by partition chromatography on Sephadex G-50 in a 2.21 x 58 cm column in the solvent system 1-butanol:pyridine:0.1% aqueous acetic acid (5:3:10) (11). The product eluted with  $R_f$  0.25 and its yield was 84 mg (81% peptide content by spectral analysis; 38% overall yield based on starting resin); tlc (1-butanol:pyridine:acetic acid:water, 5:5:1:4) on 50  $\mu$ g gave a single spot with  $R_f$  0.65 by ninhydrin and Cl<sub>2</sub>-toluidine detection; paper electrophoresis on 50  $\mu$ g samples at pH 3.7 (pyridine acetate) and pH 6.7 ( $\gamma$ -collidine acetate) gave a single spot in each case with  $R_{FYS}$  of 0.60 and 0.45, respectively (375 V, 4-5 hr, ninhydrin and Cl<sub>2</sub>-toluidine detection). Amino acid analysis of a 24-hr HCl hydrolysate gave (theoretical values in parenthesis): Lys, 4.97 (5); Asp, 1.98 (2); Thr, 2.87 (3); Ser, 1.63 (2); Glu, 3.00 (3); Pro, 1.05 (1); Gly, 3.12 (3); Ala, 1.99 (2); Val, 0.99 (1); Met, 1.04 (1); Ile, 1.34 (2); Leu, 2.06 (2); Tyr, 1.94 (2); Phe, 2.02 (2). Low value for Ile is accounted for by the acid resistant Ile-Ile sequence. Amino acid analysis of a total enzyme digest (trypsin and chymotrypsin followed by leucine aminopeptidase M) gave: Lys, 5.08 (5); Asp, 0.18 (0); Thr + Ser + Asn + Gln, 8.65 (9); Glu, 1.04 (1); Pro, 0.95 (1); Gly, 2.93 (3); Ala, 1.92 (2); Val, 1.12 (1); Met, 0.93 (1); Ile, 1.79 (2); Leu, 2.05 (2); Tyr, 2.18 (2); Phe, 2.20 (2).

The analgesic potency was estimated in mice by the tail-flick method (12) as described (2,3). The opiate receptor binding assay was carried out with rat brain membrane preparation (13) or neuroblastoma x glioma hybrid cell NG108-15 (14) as described using [<sup>3</sup>H<sub>2</sub>-Tyr<sup>27</sup>]- $\beta_h$ -EP (15) as the primary ligand and synthetic  $\beta_h$ -EP (6) as standard competing ligand. Immunoreactivity was assessed by radioimmunoassay using the published procedure (16,17).

#### RESULTS AND DISCUSSION

Biological activity of [Gln<sup>8</sup>]- $\beta_h$ -EP as assayed by various procedures is summarized in Table 1. Figure 2 presents the analgesic potency of the analog as assayed by the tail-flick test in mice. In comparison with the parent molecule replace-

Table 1

Biological Activity of Synthetic [Gln<sup>8</sup>]- $\beta$ <sub>h</sub>-Endorphin

	Opiate receptor-binding activity		Analgesic potency		Immunoreactivity	
	IC <sub>50</sub> <sup>a</sup>	Relative Potency	AD <sub>50</sub> <sup>b</sup>	Relative Potency	IC <sub>50</sub> <sup>c</sup>	Relative Potency
$\beta$ <sub>h</sub> -Endorphin	0.22 ± 0.05 <sup>d</sup>	1.0	0.098 (0.058-0.170) <sup>f</sup>	1.0	8.0 ± 0.7	1.0
	0.69 ± 0.07 <sup>e</sup>	1.0	0.028 (0.020-0.039) <sup>g</sup>	1.0		
[Gln <sup>8</sup> ]- $\beta$ <sub>h</sub> -EP	0.13 ± 0.03 <sup>d</sup>	1.7	0.036 (0.020-0.052) <sup>f</sup>	2.7	14.3 ± 0.7	0.6
	0.21 ± 0.02 <sup>e</sup>	3.3	0.013 (0.009-0.018) <sup>g</sup>	2.2		

<sup>a</sup>IC<sub>50</sub> in nM ± standard error

<sup>b</sup>AD<sub>50</sub> in nM ± (95% confidence limit)

<sup>c</sup>IC<sub>50</sub> in fM ± standard error

<sup>d</sup>Assayed with rat brain membrane preparation

<sup>e</sup>Assayed with neuroblastoma x glioma hybrid cells

<sup>f</sup>Carried out in Wisconsin

<sup>g</sup>Carried out in California

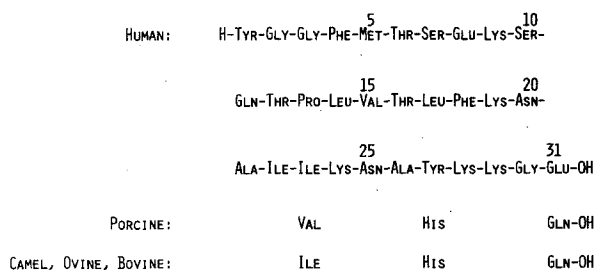


Figure 1: Amino acid sequence of human, porcine, camel, ovine, and bovine  $\beta$ -endorphin.

ment of Glu<sup>8</sup> by glutamine caused an increase of 2.2-2.7 fold analgesic potency and 1.7-3.3 fold of opiate receptor-binding activity but a loss of 40% immunoreactivity. Earlier studies (18) have shown that substitution of Glu<sup>8</sup> by glutamine elevated the immunoreactivity and opiate receptor-binding activity of  $\beta_h$ -EP-(1-27) to values nearly as great as  $\beta_h$ -EP but the analgesic potency of [Gln<sup>8</sup>]- $\beta_h$ -EP-(1-27) increased from 2% to only 12% in comparison with the potency of  $\beta_h$ -EP. In addition, replacement of Glu<sup>8</sup> by glutamine increases the opiate receptor-

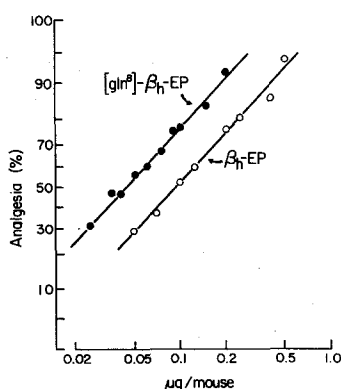


Figure 2: Comparison of antinociceptive effects of  $\beta_h$ -EP (○) and [Gln<sup>8</sup>]- $\beta_h$ -EP (●) in tail-flick test; groups of mice, 10-13 in each, were injected intracerebroventricularly. Percent analgesia as function of dose.

binding activity of synthetic [Gly<sup>31</sup>]- $\beta_h$ -EP-Gly-Gly-NH<sub>2</sub> (19) but reduces analgesic potency (20).

There are 2 Glu residues at positions 8 and 31 in the  $\beta_h$ -EP structure (Fig. 1). The analgesic potency of  $\beta_h$ -EP appears to be unchanged by the replacement of Glu<sup>31</sup> by Gly (21). In addition, camel  $\beta$ -EP with glutamine in position 31 (1; see Fig. 1) exhibits nearly identical analgesic potency (20). These observations, together with the data herein reported, suggest that glutamic acid in position 8 plays an important role in both opiate receptor-binding activity and analgesic potency of the opioid peptide. The data in Table 1 also confirm the lack of correlation between immunoreactivity and opiate receptor-binding activity and analgesic potency as reported previously (22).

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#### REFERENCES

1. Li, C. H. and Chung, D. (1976) Proc. Natl. Acad. Sci. USA, 73, 1145-1148.
2. Loh, H. H., Tseng, L-F., Wei, E., and Li, C. H. (1976) Proc. Natl. Acad. Sci. USA 73, 2895-2898.
3. Tseng, L-F., Loh, H. H., and Li, C. H. (1976) Nature 263, 239-240.
4. Li, C. H., Yamashiro, D., Tseng, L-F., Chang, W-C., and Ferrara, P. (1979) Proc. Natl. Acad. Sci. USA 76, 3276-3278.
5. Merrifield, R. B. (1964) Biochemistry 3, 1385-1390.
6. Li, C. H., Yamashiro, D., Tseng, L-F., and Loh, H. H. (1977) J. Med. Chem. 20, 325-328.
7. Erickson, R. B. and Merrifield, R. B. (1973) J. Amer. Chem. Soc. 95, 3757-3763.
8. Yamashiro, D. and Li, C. H. (1973) J. Org. Chem. 38, 591-592.
9. Yamashiro, D., Tseng, L-F., Doneen, B. A., Loh, H. H., and Li, C. H. (1977) Int. J. Peptide Protein Res. 10, 159-166.
10. Sakakibara, S., Shimonishi, Y., Kishida, Y., Okada, M., and Sugihara, M. (1967) Bull. Chem. Soc. Japan 40, 2164-2167.

11. Yamashiro, D. (1980) in "Hormonal Proteins and Peptides IX" (Li, C. H., ed.) pp. 25-107, Academic Press, New York.
12. D'Amour, F. E. and Smith, D. L. (1941) J. Pharmacol. Exp. Ther. 72, 74-79.
13. Ferrara, P., Houghten, R., and Li, C. H. (1979) Biochem. Biophys. Res. Commun. 89, 786-792.
14. Hammonds, Jr., R. G., Ferrara, P., and Li, C. H. (1981) Proc. Natl. Acad. Sci. USA 78, in press.
15. Houghten, R. A. and Li, C. H. (1978) Int. J. Peptide Protein Res. 12, 325-326.
16. Li, C. H., Rao, A. J., Doneen, B. A., and Yamashiro, D. (1977) Biochem. Biophys. Res. Commun. 75, 576-580.
17. Chang, W-C., Yeung, H. W., and Li, C. H. (1979) Int. J. Peptide Protein Res. 13, 278-281.
18. Li, C. H., Tseng, L-F., Jibson, M. D., Hammonds, Jr., R. G., Yamashiro, D., and Zaoral, M. (1980) Biochem. Biophys. Res. Commun. 97, 932-938.
19. Yamashiro, D., Ferrara, P., and Li, C. H. (1980) Int. J. Peptide Protein Res. 16, 70-74.
20. Li, C. H., Tseng, L-F., Ferrara, P., and Yamashiro, D. (1980) Proc. Natl. Acad. Sci. USA 77, 2303-2304.
21. Li, C. H., Yamashiro, D., Tseng, L-F., Chang, W-C., and Ferrara, P. (1979) Proc. Natl. Acad. Sci. USA 76, 3276-3278.
22. Li, C. H., Yamashiro, D., Tseng, L-F., Chang, W-C., and Ferrara, P. (1980) Proc. Natl. Acad. Sci. USA 77, 3211-3214.